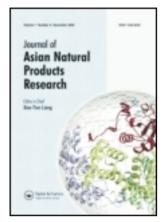
This article was downloaded by: [Malmo Hogskola]

On: 20 December 2011, At: 23:14

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered

office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information:

http://www.tandfonline.com/loi/ganp20

New chromone derivative terminalianone from African plant Terminalia brownii Fresen (Combretaceae) in Tanzania

Hiroko Negishi ^a , Takashi Maoka ^b , Marina Njelekela ^c , Naomi Yasui ^d , Sachiko Juman ^d , Jacob Mtabaji ^e , Tomohiro Miki ^d , Yasuo Nara ^f , Yukio Yamori ^d & Katsumi Ikeda ^d

Available online: 15 Mar 2011

To cite this article: Hiroko Negishi, Takashi Maoka, Marina Njelekela, Naomi Yasui, Sachiko Juman, Jacob Mtabaji, Tomohiro Miki, Yasuo Nara, Yukio Yamori & Katsumi Ikeda (2011): New chromone derivative terminalianone from African plant Terminalia brownii Fresen (Combretaceae) in Tanzania, Journal of Asian Natural Products Research, 13:03, 281-283

To link to this article: http://dx.doi.org/10.1080/10286020.2011.552431

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.tandfonline.com/page/terms-and-conditions

^a Faculty of Human Life and Environment, Nara Women's University, Nara, 630-8506, Japan

^b Research Institute for Production Development, Kyoto, 606-0805, Japan

^c Muhimbili University College of Health Sciences, Dar es Salaam, 65001, Tanzania

^d Mukogawa Women's University, Nishinomiya, 663-8179, Japan

^e Weill Bugando University College of Health Sciences, Mwanza, 1464, Tanzania

f School of Pharmacy, Shujitu University, Okayama, 703-8516, Japan

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



New chromone derivative terminalianone from African plant Terminalia brownii Fresen (Combretaceae) in Tanzania

Hiroko Negishi^a*, Takashi Maoka^b, Marina Njelekela^c, Naomi Yasui^d, Sachiko Juman^d, Jacob Mtabaji^e, Tomohiro Miki^d, Yasuo Nara^f, Yukio Yamori^d and Katsumi Ikeda^d

^aFaculty of Human Life and Environment, Nara Women's University, Nara 630-8506, Japan; ^bResearch Institute for Production Development, Kyoto 606-0805, Japan; ^cMuhimbili University College of Health Sciences, Dar es Salaam 65001, Tanzania; ^dMukogawa Women's University, Nishinomiya 663-8179, Japan; ^cWeill Bugando University College of Health Sciences, Mwanza 1464, Tanzania; ^fSchool of Pharmacy, Shujitu University, Okayama 703-8516, Japan

(Received 20 April 2010; final version received 5 January 2011)

A new chromone derivative named terminalianone (1) was isolated from the African plant, *Terminalia brownii* Fresen (Combretaceae) in Tanzania. Its structure was determined to be 7-hydroxy-3-[6'-hydroxyphenyl-2'-oxo-ethyl]chromone by FAB-MS and NMR spectral data.

Keywords: *Terminalia brownii* Fresen; Combretaceae; chromone; 7-hydroxy-3-[6'-hydroxyphenyl-2'-oxo-ethyl]chromone

1. Introduction

The African plant, Terminalia brownii Fresen (Combretaceae), is found in many regions of Africa. It has different vernacular names in different places such as 'orbukoi' in Maasai of Tanzania and kuuku, muvuku in Kamba of Kenya. Traditional healers in Africa have used Combretum and Terminalia species due to their antiinfective effect, including antibacterial, antifungal, and antiparasitic activities [1-4]. The Combretaceae family is the source of a wide range of tannins, flavonoids, terpenoids, and stilbenoids. It was reported that the leaves, barks, and fruits of Terminalia arjuna, Terminalia bellerica, Terminalia chebula, and Terminalia muelleri; the leaves and fruits of Phyllanthus emblica; and the seeds of Syzygium cumini were found to have high total phenolic contents and high antioxidant activity [5]. Various types of flavonoids detected from the members of the Combretaceae family include arjunolone, flavones, bicalein, quercetin, kempferol, pelorgonigin, and resveratrol-3-O- β -D-rutinoside [6–9].

In this study, a new chromone was isolated from *T. Brownii* Fresen in Tanzania, and its structural elucidation is described (Figure 1).

2. Results and discussion

Preparative HPLC of polyphenols on octa decyl silyl (ODS) with 33% $\rm CH_3CN-H_2O$ afforded a new compound (1). Compound 1 showed a quasi-molecular ion at m/z 297.0766 [M + H]⁺ compatible with the formula of $\rm C_{17}H_{13}O_5$ by positive ion high-resolution FAB-MS. Thus, the molecular formula of 1 was deduced to be $\rm C_{17}H_{12}O_5$. The acetylation of compound 1 provided a diacetate derivative and indicated the presence of two hydroxyl groups in the molecule. The $\rm ^1H$ NMR spectral data for 1

^{*}Corresponding author. Email: negishi@cc.nara-wu.ac.jp

Figure 1. Structure of compound 1.

showed two methylene protons and eight methine protons, including one singlet, six doublet, and one double doublet signals (Table 1). The 1 H NMR signals at δ 6.88 (2H, d, J = 9.0 Hz) and 7.99 (2H, d, $J = 9.0 \,\mathrm{Hz}$) were assigned to an AA'BB' spin system of a 1,4-disubstituted phenyl group. The signals of ABX spin system at δ 6.85 (1H, d, $J = 2.5 \,\text{Hz}$), 6.92 (1H, dd, J = 8.5, 2.5 Hz), and 7.91 (1H, d, $J = 8.5 \,\mathrm{Hz}$) indicated the presence of a 1,3,4-trisubstituted phenyl group. They were confirmed by ¹H-¹H COSY experiment (Figure 2). The ¹³C NMR and DEPT experiments revealed one sp³ methylene, eight methines, and eight quaternary carbons, including two hydroxyl substituted carbons (δ 164.0 and 164.6) and two carbonyl carbons (δ 178.8 and 197.4) as shown in Table 1. The complete assign-

Table 1. ¹H (500 MHz) and ¹³C (125 MHz) NMR spectral data for **1** in CD₃OD.

Position	$\delta_{ m C}$	$\delta_{ m H}$	
2	156.1	8.05	S
3	120.2		
4	178.8		
5	128.1	7.91	d (8.5)
6	116.3	6.92	dd (8.5, 2.5)
7	164.0		
8	103.4	6.85	d (2.5)
9	160.3		
10	117.6		
1'	35.4	4.10 (2H)	S
2'	197.4		
3'	129.8		
4' 5'	132.1	7.99	d (9.0)
5'	116.3	6.88	d (9.0)
6'	164.6		
7'	116.3	6.88	d (9.0)
8'	132.1	7.99	d (9.0)

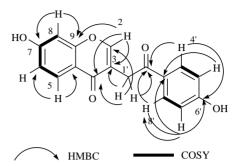


Figure 2. Key HMBC and COSY correlations of compound 1.

ments of direct ¹H-¹³C connections were established by HSQC experiment. The long-range ¹H-¹³C correlations were analyzed by HMBC experiment. The HMBC correlations from H-2 (δ 8.05) to C-3 (δ 120.2), C-4 (δ 178.8), and C-9 (δ 160.3); from H-8 (δ 6.85) to C-7 (δ 164.0) and C-9 $(\delta \ 160.3)$; from H-5 $(\delta \ 7.91)$ to C-10 $(\delta \ 160.3)$ 117.6) and C-6 (δ 116.3) revealed the presence of a 3-substituted-7-hydroxychromone moiety in 1 (Figure 2). The remaining structural moiety was elucidated to be a para-hydroxyphenyl-2-oxo-ethyl by HMBC correlations. The HMBC correlations from H-1' (δ 4.10) to C-2' (δ 197.4), from H-4' and H-8' (δ 7.99) to C-2' (δ 197.4) and C-6' (δ 164.6), from H-5' and H-7' (δ 6.88) to C-3' (δ 129.8) were consistent with a para-hydroxyphenyl-2-oxo-ethyl moiety (Figure 2). Furthermore, the HMBC correlations from H-1' (δ 4.10) to C-2 (δ 156.1), C-3 (δ 120.2), and C-4 $(\delta 178.8)$ revealed that this structural moiety was substituted at the C-3 position of 7-hydroxy-chromone ring as shown in Figure 1. Therefore, the structure of 1 was determined to be 7-hydroxy-3-[6'-hydroxyphenyl-2'-oxo-ethyl]chromone, terminalianone.

3. Experimental

3.1 General experimental procedures

The positive ion FAB-MS spectra were recorded using a JEOL JMS-HX 110A

mass spectrometer with m-nitrobenzyl alcohol as a matrix. The 1 H NMR (500 MHz) and 13 C NMR (125 MHz) spectra were measured with a Varian UNITY INOVA 500 spectrometer in CD₃OD with TMS as an internal standard. Preparative HPLC was performed on a Shimadzu LC-6AD with a Shimadzu SPD-6AV spectrophotometer set at 280 nm. The column used was a 250 \times 10 mm i.d., 10- μ m LiChrospher RP-18 (e) (Cica-Merck, Darmstadt, Germany).

3.2 Plant material

The barks of *T. brownii* Fresen were collected by Maasai in Arusha, Tanzania, in July 2005. A voucher specimen (2005TZ01) has been deposited at Mukogawa Women's University, Nishinomia, Japan.

3.3 Extraction and isolation

The air-dried and powdered plants (6.94 g) were extracted with 100 ml of water at 100°C for 15 min. The extract was centrifuged at 3000 rpm/min for 5 min, and the supernatant was collected. Polyphenols were separated by preparative HPLC on LiChrospher RP-18 with 33% CH₃CN-H₂O at a follow rate of 2.0 ml. Compound 1 (5 mg) was obtained from the peak at a retention time of 22 min.

3.3.1 *Compound* **1**

Compound 1, powder was UV (MeOH) λ_{max} : 280 nm. ¹H and ¹³C NMR spectral data: see Table 1. HR-ESI-MS: m/z

297.0766 $[M + H]^+$ (calcd for $C_{17}H_{13}O_5$, 297.0759).

3.4 Acetylation of 1

About 0.5 mg of 1 was dissolved in 0.5 ml of dry pyridine. Then, 0.5 ml of acetic anhydride was added to this solution and the solution was allowed to stand for 5 h at room temperature. After that, the reaction product was extracted with ether by adding water.

Diacetate of 1. Positive ion FAB-MS: m/z 403.0800 $[M + Na]^+$ (calcd for $C_{21}H_{16}O_7Na$, 403.0793).

References

- [1] Z.H. Mbwambo, M.J. Moshi, P.J. Mashimba, M.C. Kapingu, and R.S.O. Nondo, BMC Complement Altern Med. 7 (2007), doi: 10.1186/1472-6882-7-9.
- [2] J.N. Eloff, D.R. Katerere, and L.J. McGaw, J. Ethnopharmacol. 119, 689 (2008).
- [3] P. Fyhrquist, L. Mwasumbi, C.A. Hæggström, H. Vuorela, R. Hiltunen, and P. Vuorela, J. Ethnopharmacol. 79, 169 (2002).
- [4] L.G. Chen, W.T. Huang, L.T. Lee, and C.C. Wang, *Toxicol. In Vitro* 23, 603 (2009).
- [5] M. Bajpai, A. Pande, S.K. Tewari, and D. Prakash, *Int. J. Food Sci. Nutr.* **56**, 287 (2005).
- [6] G.R. Pettit, M.S. Hoard, D.L. Doubek, J.M. Schmidt, R.K. Pettit, L.P. Tackett, and J.C. Chapuis, J. Ethnopharmacol. 53, 57 (1996).
- [7] D.S. Kumar and Y.S. Prabhakar, *J. Ethnopharmacol.* **20**, 173 (1987).
- [8] D.R. Katerere, A.I. Gray, A.R. Nash, and R.D. Waigh, *Phytochemistry* 65, 433 (2004).
- [9] S. Dwivedi, J. Ethnopharmacol. **114**, 114 (2007).